# Job aid BD FACSDiva<sup>™</sup> Software Determining initial PMT voltages

This job aid contains instructions for determining voltage settings that will optimize the stain index (SI) of cells stained with a reference reagent for each detector. After PMT voltages have been determined, they can be saved as Application Settings and used as a starting point for subsequent experiments.

# Before you begin

- Prepare cells; for this method only unstained processed cells are required, no single-stained cells are needed.
- Prepare a tube with 1 drop of mid-peak Sphero<sup>®</sup> Rainbow Fluorescent Particles (Cat. No. 556298) to be used as the
  postive control.
- Ensure that the instrument has been started up and a CS&T performance check has been performed.

#### Setting up an experiment

Select Edit > User Preferences and set the following:

- Under the **General** tab, clear the checkbox for **Remove** tube-specific cytometer settings on duplicate.
- Clear the checkbox for **Save analysis after recording** through a global worksheet.
- 2 Select File > Import > Experiments and select A5 SE Voltage Titration Template experiment. This template is only compatible with the BD FACSymphony" A5 SE Cell Analyzer's default CST configuration, running BD FACSDiva" Software version 9.3 or later. Skip to the section titled Recording Data Files after importing the BD FACSDiva" Software template. If you'd like to create your own experiment template then proceed to step 3.
- 3 Create and rename a new experiment.
- Select the experiment in the Browser and then in the Inspector, clear the checkbox for Use global cytometer settings.
- 5 Add a specimen and rename it **Unstained Cells**.
- 6 Select Tube\_001 and click the New Cytometer Settings button to add tube-specific cytometer settings to Tube\_001.
- 7 Set PMT voltages for each fluorescence detector to 200.

User Preferences						
General Gates Worksheet Plot FCS	5 Templates Statistics Biexponential					
Tube-specific worksheet						
Start acquisition on pointer chang	je					
Show file identifier (GUID) in stati	istics view					
Remove tube-specific cytometer s	settings on duplicate					
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Show display of size column in the Experiment browser						
A5 SE Voltage Titration Template     Cytometer Settings     Global Worksheets     Mustained Cells     Tube_001	Inspector - A5 SE Voltage Titration Template     Experiment Keywords     Name: A5 SE Voltage Titration Template     Owner: Administrator     Modified: 8/23/21, 7:50:32 AM     Config: FACSymphony A5 SE Default Configuration     Log Decades for Plots     ④ 4 Log Decades     ⑤ 5 Log Decades     ① Use global cytometer settings					
🖻 Browser - A5 SE Voltage Titration Template						
A5 SE Voltage Titration Template  Cytometer Settings  Global Worksheets  Mustained Cells						

🖨 🚺 Tube\_001

😝 Cytometer Settings

- 8 On the global worksheet, create an FSC vs. SSC dot plot.
- 9 Create histograms for each detector on your instrument.
- 0 Create a P1 gate on the FSC vs. SSC plot and set all histograms to show P1.

#### Setting voltages

- Load a tube of unstained cells on the cytometer and start acquiring.
- 2 Adjust FSC and SSC voltages to place the cells on scale and set FSC threshold, if needed.
- 3 Load a tube of the mid-peak beads and confirm they are still on scale; adjust FSC and SSC voltage, if needed
- 4 Adjust P1 to identify the population of interest.
- 5 In the **Experiment Layout**, **Acquisition** tab, select your global worksheet from the **Global Worksheet** menu.
- 6 Select **P1** as the **Stopping Gate**.
- Enter 2,000 in the Events to Record field.
- 8 Duplicate **Tube\_001** and increase the PMT voltages for each fluorescence detector by 20 volts.
- 9 Repeat step 8 to create tubes for voltages up to 800 volts, for a total of 31 tubes.
- 0 Duplicate the specimen and rename it **Beads**.

### Recording data files

- Acquire the unstained cells and record a data file for Tube\_001 in the Unstained Cells specimen.
- 2 Record data files for all tubes in the specimen using the same sample.
  - Keep the same flow rate for all tubes.
  - Record data for all 31 tubes.

3 Repeat steps 1 and 2 with the beads.



<b>2</b>	Experiment Layout							
La	abels	Keywords A	cquisition					
	Qu Ev Gle	ick Entry ents to Record obal Worksheet	10,000 ~	Stopping G Storage Ga	ate p1 te	<ul><li>✓ Stopping Tim</li><li>✓</li></ul>	e (sec)	0
	Name			Events to Rec	Global Worksh	Stopping Gate	Storage Gate	
	٠	PMT V	oltration A5 SE Exp	periment				
	•	<ul> <li>Unstained Cells</li> </ul>				Global Sheet1		
	r Tube_001			2,000		P1 (GW) 🗸	All Events (G	

#### Status Parameters Threshold Laser Compensation

	Parameter	Voltage				
	FSC	475				
	SSC	325				
# U379	UV379	320				
# U446	UV446	320				
+ U515	UV515	320				
+ U540	UV540	320				
# U585	UV585	320				
# U610	UV610	320				
# U660	UV660	320				
# U695	UV695	320				
# U736	UV736	320				
+ U809	UV809	320				
<ul> <li>V427</li> </ul>	V427	320				

## Analyzing data

• For **Tube\_001** of the unstained cell specimen, create a statistics view to display median and rSD for all the fluorescent detectors.

2 Right-click the specimen and select **Batch Analysis**.

3 Make the following selections in the **Batch Analysis** dialog:

- Select **Auto** and set to 0 seconds.
- Select **Statistics**.
- For **Stats Filename**, select a file location for the CSV file and rename it **Unstained Cells**.
- Deselect all other check boxes
- 4 Edit the statistics view and deselect the **rSD column**. Repeat steps 2 and 3 with the **Beads Specimen**.

Calculate the stain index for each PMT voltage data point using the A5 SE Voltage Titration Worksheet Template provided by BD. Open the CSV file from the Beads Specimen batch analysis and use shift to select and copy all cells containing values, excluding the top header row. Paste into the highlighted area in the Positives tab of the PMTV Setup Template.

 Repeat step 5 for the Unstained Cells specimen using the Negatives tab in the A5 SE Voltage Titration Worksheet Template.

Identify PMT voltages where the stain index for a biological sample is at or near the maximum voltage for each detector.

If you need assistance identifying the optimal PMT voltages or have questions about the PMTV Setup Template document, contact **ResearchApplications@bd.com**.

Edit Statistics View												
Header Populations	Statistics											
Parameters		Min	Max	Geo	Mean	Median	50	50	□ %CV	2%rCV	Mode	
• FSC-A												
<ul> <li>SSC-A</li> </ul>												1
• Time												1
<ul> <li>UV379-A</li> </ul>						$\checkmark$		$\checkmark$				1
<ul> <li>UV446-A</li> </ul>												1

🛃 Batch Analysis	s
Auto	🗌 Output To Printer 🛛 🖓 Statistics
View Time:	0 🗸 🗌 Save as PDF 🔄 Freeze Biexponential Scales
Manual	Save as XML Use Preferred Global Worksheet
	✓ Add Report
PDF Filename:	riment-Batch_Analysis_11022022110111.pdf Browse View PDF
XML Filename:	riment-Batch_Analysis_11022022110111.xml Browse
Stats Filename:	riment-Batch_Analysis_11022022110111.csv Browse
Status:	0%
	Start Pause Continue Close

# Saving as application settings

- 1 Create a new experiment in the Browser and enter the voltages determined in the previous section.
- 2 Right-click the cytometer settings and select **Application Settings** > **Save**.
  - The application settings are saved in the Cytometer Catalog and can be applied to future experiments with the same configuration.
  - Application settings will be updated automatically when a performance check has been run.



BD flow cytometers are Class 1 Laser Products. For Research Use Only. Not for use in diagnostic or therapeutic procedures.

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