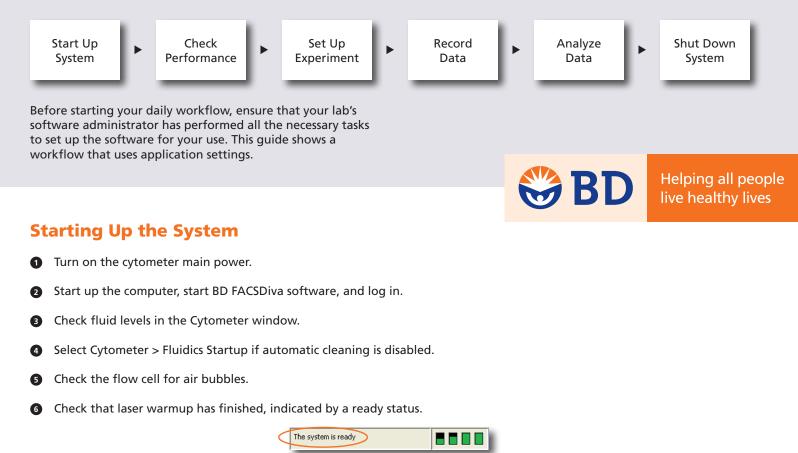
BD FACSDiva Software Quick Reference Guide for BD FACSCanto Systems

This guide contains instructions for using BD FACSDiva[™] software version 6.0 and later with BD FACSCanto[™] and BD FACSCanto II systems.

Workflow Overview

The following figure shows the steps for daily workflow using BD FACSDiva software.



Checking Cytometer Performance

1 Select Cytometer > CST.

	🕅 Cytometer Setup and Tracking									
	Ele Cytometer Iools									
	Setur Reports Performance Tracking									
	System Summary: OK	Setup Control - Research Use Only Load a tube with beads and click Run button to start Check Performance.								
Verify the Cytomter Configuration and	Cytometer Configuration: 2-laser, 6-color (4-2) (8D default) Lot ID: 68341	Run Abort Load Tube Manually								
bead Lot ID.	Cytometer Baseline: May 23, 2007 12:27 PM Cytometer Performance: July 13, 2007 02:48 PM Cytometer Performance Results: Passed	Cytometer Configuration 2-4see, 6-color (+2) (8D default) Select Configuration Setup Beads Let ID: 663911	If needed, select a different configuration							
	Usulieter Ferdillerite Freudust, Fasser	Product: CST Setup Beads Part II: 633391 Expiration Date: 10-10-2007	or bead lot ID.							

- 2 Run the BD[™] Cytometer Setup and Tracking beads.
- **3** View the Cytometer Performance Report.
- Close the Cytometer Setup and Tracking window.

Setting Up the Experiment

- Select Edit > User Preferences and verify that selected preferences are appropriate.
- 2 Create an experiment in the Browser.
- 8 Right-click Structure Settings in the Browser. Select Application Settings > Apply.

See page 5 for additional information about creating application settings.

	Application Settings		Cytometer Settings Mismatch			
	Cytometer Configuration: 2-laser, 7-c	olor (5-2) (BD default) 🛛 👻	The application settings to be applied do not match the selected cytometer settings. The following parameters are not in the cytometer settings to be applied: PE-Cy7-A.			
Select an application setting.	Name > Owner Application A UserName1 Application B UserName1	Date Created 07/16/07 11:39:58 AM 07/16/07 11:40:06 AM	Click Apply to apply PMT Volkage and Threshold values only for matching parameters.			
	View	Apply Cancel	Click Overwrite to replace all parameters and values with those from the selected application settings.			
			Apply Overwrite Cancel			

• Select Experiment > Compensation Setup > Create Compensation Controls.

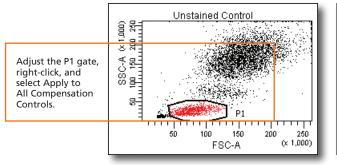
Create Compensation Com Tut Tut Tutude separate unstained co	pes Plate	
Fluorophore	Label	
FITC	Generic	
• PE	Generic	
PerCP-Cy5-5	Generic	Create label-specific
• PE-Cy7	Generic	controls as needed.
# APC	Generic	
# APC-Cy7	Generic	
Add Delete	Labels OK Cancel	

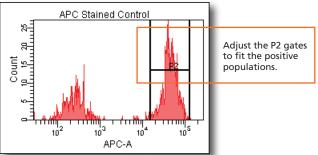
Install the unstained control tube onto the cytometer. Click Acquire Data

	🕷 Cytometer - FACSCanto (V0041) 🛛 🔀	🛗 Normal Worksheet - Unstained Control	
Verify that the FSC, SSC, and threshold settings are appropriate.	Laser Compensation Ratio Status Parameters Threshold	Image: Sheet1 Image: Sheet1<	View data in the normal worksheets provided.
	Parameter Voltage	Galactic Control	
	FITC 530 V _ PE 473 V _ PerCP-Cy5-5 637 V _		
	PE-Cy7 778 V APC 613 V APC-Cy7 641 V V V		
		FOC-A CONTRACT	

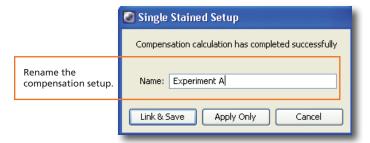
6 Record data for the compensation control tubes.

View the recorded data and gate the positive populations.



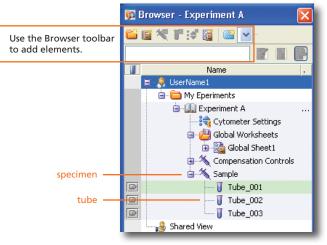


3 Select Experiment > Compensation Setup > Calculate Compensation.

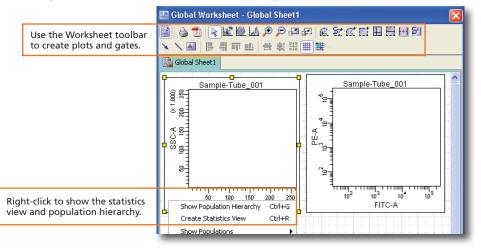


Recording Specimen Data

1 Create Browser elements.



2 Create plots, gates, and statistics needed for recording.



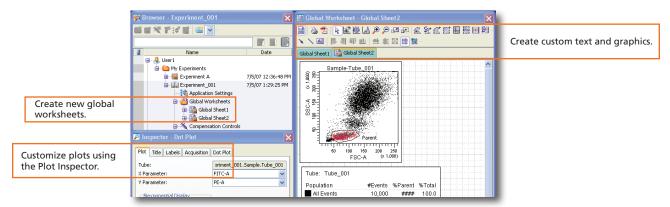
3 Make entries in the Experiment Layout.

	C Experiment Layout											
Specify reagent labels, keywords, and acquistion criteria as needed.	Lai	abels / Keywords / Acquisition							Labels Name			
		_	Name		Label	Label	Label	Label	Label	Label		List by user UserName1
		•	🔨 Sample								^	🗄 🛞 BD Defined
			🗍 Tube_001		FITC CD3	PE	PerCP-Cy5-5	PE-Cy7	APC	APC-Cy7		Add to List Delete from List
		•	🛛 🗍 Tube_002		FITC	PE	PerCP-Cy5-5	PE-Cy7	APC	APC-Cy7		Assign or Remove Labels
		•	↓ Tube_003		FITC	PE	PerCP-Cy5-5	PE-Cy7	APC	APC+Cy7	×	Assign Remove
												OK Cancel

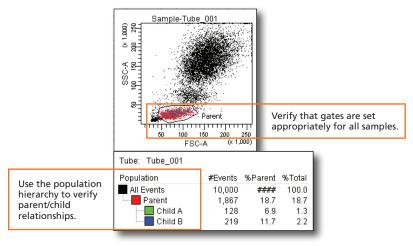
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Analyzing Data

1 Create plots, gates, and statistics needed for analysis.



2 Perform quality control of the analysis.



- 3 Do one of the following to print or export the results.
 - Select File > Print to print the active worksheet.
 - Select File > Export to export selected elements.
 - Right-click a specimen or experiment and select Batch Analysis (using a global worksheet).

	💽 Batch Analys		
	 O Auto View Time: 1 ○ Manual 	♥ Output To Printer ♥ Statistics ● ♥ Save as PDF ♥ Freeze Biexponential Scales ♥ Add Report to PDF ♥ Use Preferred Global Worksheet	Select to print, save as a PDF, or export the statistics as needed.
Specify where to save the PDF and exported statistics files.	PDF Filename: Export Filename:		
	Status:		

4

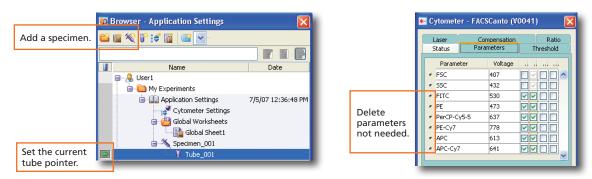
Shutting Down the System

- 1 Perform a fluidics shutdown.
- 2 Empty the waste and refill fluids if prompted to do so.
- 3 Turn off the cytometer main power and shut down the computer.

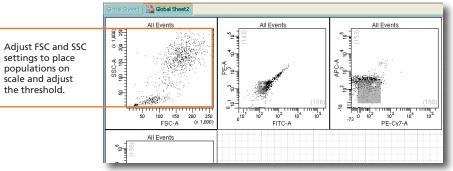
Creating Application Settings

Before creating application settings, perform the cytometer startup procedure according to your cytometer user's guide and run a performance check.

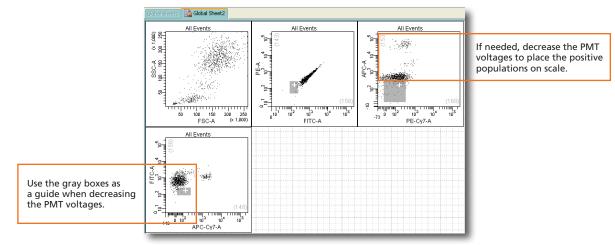
1 Create a new experiment.



- 2 Right-click Cytometer Settings in the Browser. Select Application Settings > Create Worksheet.
- Install the unstained control onto the cytometer. Click
- Adjust the cytometer settings.



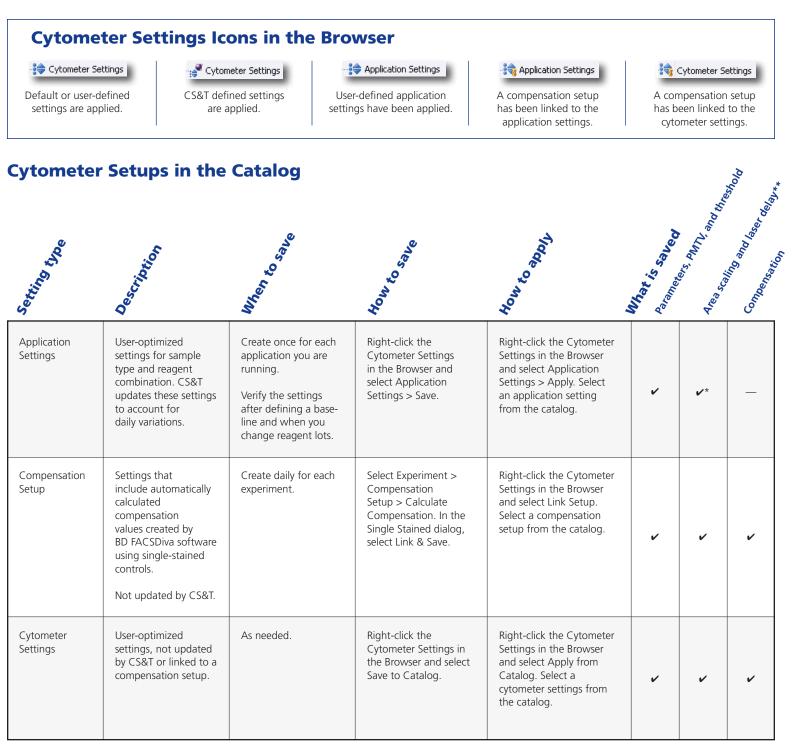
6 Acquire single-stained controls.



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6 Right-click 😫 Cytometer Settings in the Browser. Select Application Settings > Save.





* The area scaling is adjusted for the CS&T beads only. You may need to readjust this setting for your cell type.

** The latest optimized laser delay setting is always used on the cytometer. When the saved cytometer settings are re-applied, the laser delay does not change to the value saved with the settings. The laser delay setting is saved for reference only.

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