

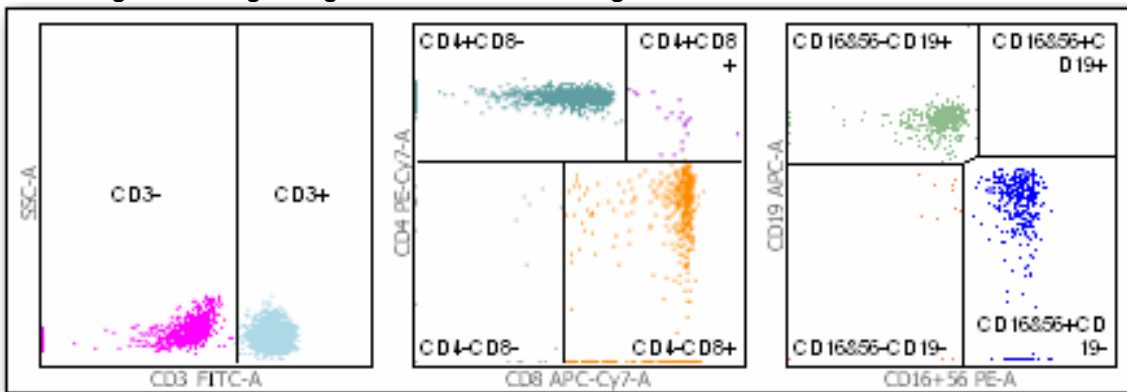
Troubleshooting Results with BD FACSCanto™ Clinical Software

Interpreting plots with dots on or near the axes

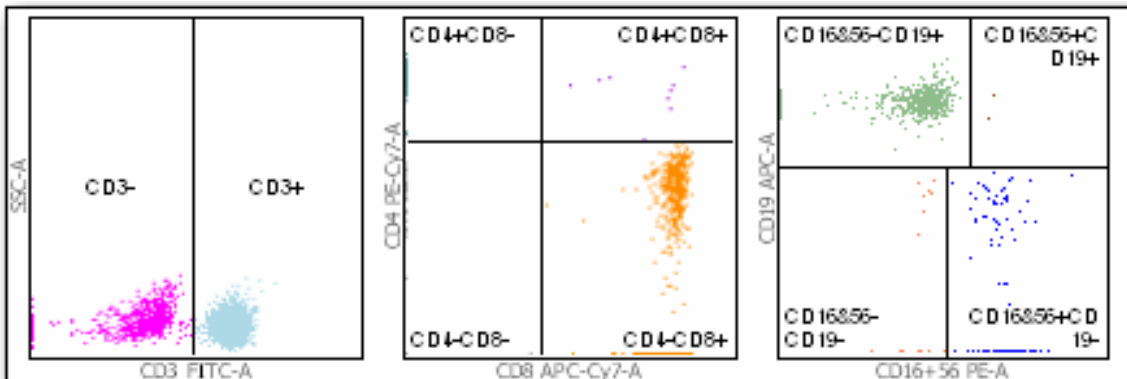
Immunofluorescence measurements in flow cytometry might result in signals that measure at or very near zero (mV). Since $\text{Log}(0)$ is undefined, these events are assigned to the baseline (axis) bin of the log display scale. Similarly, logarithm values with low signal intensities that result in negative values are also assigned to this baseline bin. See the following figure (top row). Under rare conditions, events from a given patient sample may be predominantly assigned to this baseline bin due to very low signals. See the following figure (bottom row).

However, the higher signal intensities from the population will still be on scale. All of the binned events, including the baseline and above, are part of the original lymphocyte and gate classification. Therefore, the baseline bin events are correctly included in the numerical results shown on the Laboratory Report. This is expected behavior and does not impact the numerical results or the performance of the assay.

Low voltages resulting in negative values can be assigned to the baseline bin





Predominantly low signals for a given patient, assigned to the baseline bin

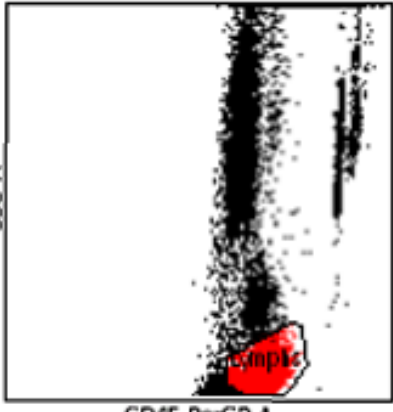
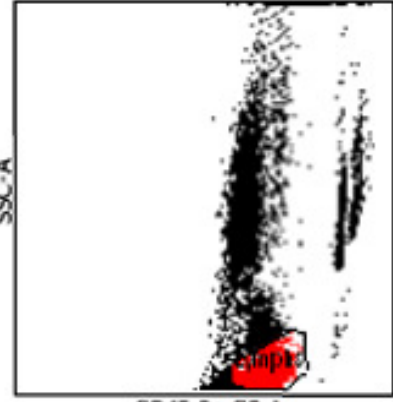


Troubleshooting Results with BD FACSCanto™ Clinical Software

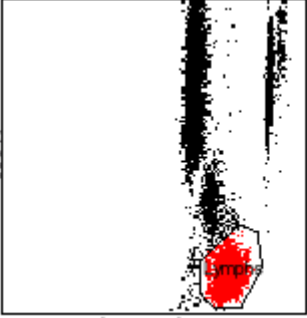
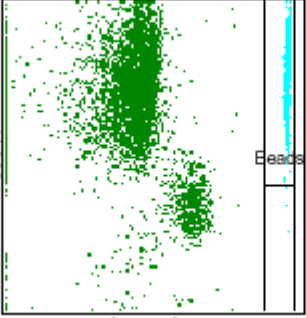
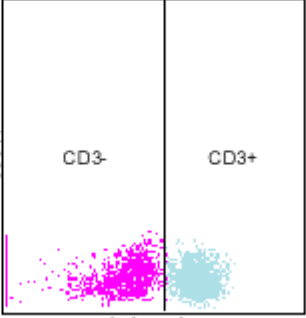
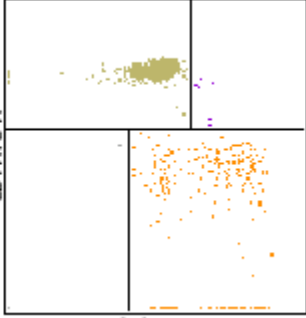
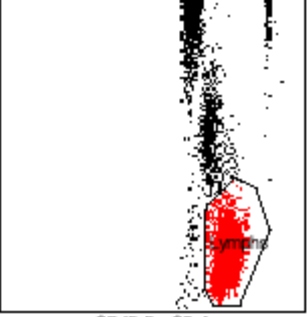
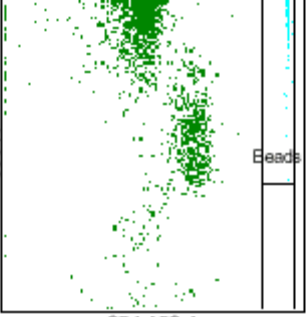
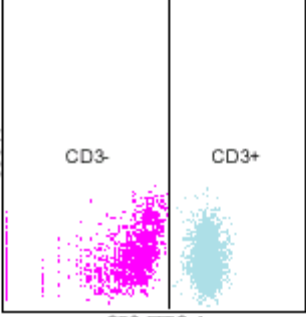
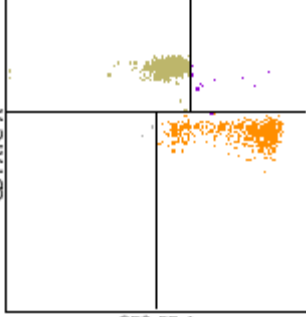
How do we analyze and gate commercial process controls with BD FACSCanto clinical software?

Observation	Possible Causes	Recommended Solutions
Cell populations in the CD45 vs SSC plot extend upward	Inadequate lysing of the sample	Prepare the sample again, and ensure that it is completely lysed.
 <p>CD45 PerCP-A</p>		
Granulocytes with low side scatter in the CD45 vs SSC plot, no distinct monocyte population	Aged blood or stained cells	See the reagent package insert for stability limitations.
 <p>CD45 PerCP-A</p>		

Troubleshooting Results with BD FACSCanto™ Clinical Software

<p>Debris encroaching on populations in the CD45 vs SSC plot</p>	<ul style="list-style-type: none"> • Excessive mixing • Aged blood or stained cells 	<p>Prepare the sample again.</p>
 <p>SSC-A</p> <p>CD45 PerCP-A</p>		
<p>Observation</p>	<p>Possible Causes</p>	<p>Recommended Solutions</p>
<p>Vertically compressed populations</p>	<p>Side scatter too low</p>	<p>Reacquire the sample. Set SSC so granulocytes reach to top of the CD45 vs SSC plot.</p>
	<p>Lipemic sample</p>	<p>Check the reagent package insert for instructions.</p>
 <p>SSC-A</p> <p>CD45 PerCP-A</p>		

Troubleshooting Results with BD FACSCanto™ Clinical Software

<p>Indistinct populations; events sparse or missing from one population; lack of separation between the CD3⁻ and CD3⁺ cluster</p>	<p>Incorrect spectral overlap</p>	<p>Re-run the setup, optimizing for the application. Re-run the sample.</p>	
 <p>CD45 PerCP-A</p>	 <p>CD4 APC-A</p>	 <p>CD3 FITC-A</p>	 <p>CD4 APC-A</p> <p>CD8 PE-A</p>
<p>Granulocytes cut off at the top of the plot; stretched monocyte population</p>	<p>High SSC</p>	<p>Re-run the setup, optimizing for the application. Re-run the sample, lowering the SSC.</p>	
 <p>CD45 PerCP-A</p>	 <p>CD4 APC-A</p>	 <p>CD3 FITC-A</p>	 <p>CD4 APC-A</p> <p>CD8 PE-A</p>