Fixable Viability Stain 660

Product Information

Material Number: 564405
Size: 100 µg

Description

BD Horizon™ Fixable Viability Stain 660 (FVS660) is useful for discrimination of viable from non-viable mammalian cells in multicolor flow cytometric applications. This dye reacts with and covalently binds to cell-surface and intracellular amines. Permeable plasma cell membranes, such as those present in necrotic cells, allow for the intracellular diffusion of the dye and covalent binding to higher overall concentrations of amines than in non-permeable live cells. Therefore, necrotic cells present in a typical in vitro assay label with higher levels of dye increasing their fluorescence intensity 10-20 fold over that of viable cells. The labeled cells can be fixed with formaldehyde for downstream decontamination, freezing and/or permeabilization and subsequent intracellular staining while maintaining stable viability stain fluorescence.

BD Horizon™ Fixable Viability Stain 660 is excited by the Red laser (with an excitation maximum of 649 nm) and has a fluorescence emission maximum of 660 nm.

Application Notes

Application

- Flow cytometry: Tested During Development
- Intracellular staining (flow cytometry): Tested During Development

Recommended Assay Procedure:

Preparation

Bring FVS660 dye powder and 190 µl of fresh cell culture-grade Dimethyl Sulfoxide (DMSO; e.g., Sigma D2650) to room temperature. Add 190 µl of DMSO and vortex solution well. Inspect the solution and repeat vortex until the stock dye has fully dissolved. This is the Stock Solution.

Storage

Upon arrival, store the dry dye desiccated and protected from light at -80°C until use. After reconstitution with DMSO, store the Stock Solution at -20°C in small aliquots. Do not use reconstituted dye after 90 days of storage. Please discard the dye solution after 90 days post reconstitution with DMSO.
Cytometry Requirements

Red laser-equipped Flow Cytometers (e.g., BD FACSCanto™ II, BD LSRRfortessa™, BD™ LSR II, or BD Accuri™ C6) can be used. This dye can be read out of filters commonly used for APC or Alexa Fluor® 647 (e.g., 660/20 or 670/30). Fluorescence compensation is best achieved using a sample of the cells of interest.

Procedure

Fixable Viability Stain 660 labeling of cells
1. Prepare cells for flow cytometric staining using sodium azide-free buffers.
2. Wash cells one time in sodium azide- and protein-free Dulbecco’s Phosphate Buffered Saline (1X DPBS).
3. Resuspend cells at 1-10x10^6 cells/ml in sodium azide- and protein-free 1X DPBS.
4. Add 1 μl of the BD Horizon™ Fixable Viability Stain 660 Stock Solution for each 1 ml of cell suspension (1:1000) and vortex immediately.
   a. Note: We recommend titrating the dye for optimal performance, as different cell types and different applications can result in a wide degree of variability in staining. Please read Note 1 below for guidance on recommended ranges.
5. Incubate the mixture for 10-15 minutes at room temperature or 2-8°C protected from light.
   a. Optional: Alternatively, incubate mixtures at 37°C for 5-7 minutes.
6. Wash cells twice with 2 ml of BD Pharmingen™ Stain Buffer (FBS) (Cat. No. 554656) or the equivalent.
7. Decant the supernatant and gently mix to disrupt the cell pellet.
8. Resuspend the cells in Stain Buffer (FBS) or equivalent.
9. Stain, fix and permeabilize cells as desired for downstream applications.

Notes:
1. Each user should determine the optimal concentrations of reagents, cells, and conditions for the assay of interest. We recommend titrating the reagent in early experiments to obtain optimal results. The following are ranges that we have found to work for various cell systems:
   a. Lysed Whole Blood: 1:1,000 from the Stock Solution.
   b. Primary Cells: 1:1,000 - 1:4,000 from the Stock Solution.
   c. Cell Lines: 1:4,000 - 1:10,000 from the Stock Solution.
2. The reactivity of the free dye is quenched by washing with buffer containing protein (e.g., FBS or BSA).
3. Cells may be stained in bulk prior to freezing or staining with fluorescent antibodies.
4. BD Horizon™ Fixable Viability Stain 660 can be used in intracellular staining assays that require fixation with formaldehyde and permeabilization with methanol and detergents such as those used for BD Phosflow™ staining (e.g., Cat. No. 558050, BD Phosflow™ Perm Buffer III), intracellular cytokine staining (e.g., Cat. No. 554714, BD Cytofix/Cytoperm™ Fixation/Permeablization Kit), or transcription factor staining (e.g., Cat. No. 562574/562725, BD Pharmingen™ Transcription Factor Buffer Set).
5. Apoptotic cells can show variable staining. We recommend co-staining with, e.g., Annexin V FITC (Cat. No. 556419) if further analysis is desired for the apoptotic cells.

Suggested Companion Products

<table>
<thead>
<tr>
<th>Catalog Number</th>
<th>Name</th>
<th>Size</th>
<th>Clone</th>
</tr>
</thead>
<tbody>
<tr>
<td>554656</td>
<td>Stain Buffer (FBS)</td>
<td>500 mL</td>
<td>(none)</td>
</tr>
<tr>
<td>554655</td>
<td>Fixation Buffer</td>
<td>100 mL</td>
<td>(none)</td>
</tr>
<tr>
<td>557885</td>
<td>Perm/Wash Buffer I</td>
<td>125 mL</td>
<td>(none)</td>
</tr>
<tr>
<td>558050</td>
<td>Perm Buffer III</td>
<td>125 mL</td>
<td>(none)</td>
</tr>
<tr>
<td>562574</td>
<td>Transcription Factor Buffer Set</td>
<td>100 Tests</td>
<td>(none)</td>
</tr>
<tr>
<td>562725</td>
<td>Transcription Factor Buffer Set</td>
<td>25 Tests</td>
<td>(none)</td>
</tr>
<tr>
<td>554714</td>
<td>BD Cytofix/Cytoperm™ Fixation/Permeablization Kit</td>
<td>250 Tests</td>
<td>(none)</td>
</tr>
<tr>
<td>554657</td>
<td>Stain Buffer (BSA)</td>
<td>500 mL</td>
<td>(none)</td>
</tr>
</tbody>
</table>

Product Notices

1. Since applications vary, each investigator should titrate the reagent to obtain optimal results.
2. For fluorochrome spectra and suitable instrument settings, please refer to our Multicolor Flow Cytometry web page at www.bdbiosciences.com/colors.
3. Alexa Fluor® is a registered trademark of Molecular Probes, Inc., Eugene, OR.
4. Before staining with this reagent, please confirm that your flow cytometer is capable of exciting the fluorochrome and discriminating the resulting fluorescence.

References


